


Use of Promega's Casework Direct Extraction Kit as a Y-Screening Tool and for the Rapid Processing of Touch DNA Samples

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Columbus Forensic Services Center


- Service the City of Columbus, Ohio
 - population approx. 1,000,000
- Forensic Biology, Firearms, Drug Chemistry, Questioned Documents, Latent Print Processing and Latent Print Comparisons
- (9) DNA analysts + DNA Technical Leader
- 2017 received 2,280 lab requests and released 2,269 reports



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Methods: Y-screening with manual differentials

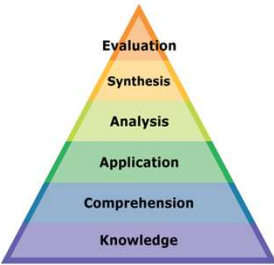
- Samples
 - Use a "Direct to DNA" approach (SAEK swabs)
 - Triage samples for 1st or 2nd round processing
- Extraction
 - Differential extraction on the Qiagen EZ1[®] robot
 - Sperm fraction (SF) spotted on slide (5µl) following last wash step
- Quantification
 - All samples using Plexor[®] HY System
- Amplification
 - Fusion 6C[®] and PowerPlex[®] Y23
- Loading
 - 3500 (STR and Y-STR) Genetic Analyzers



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Concerns

- Time (screening/lab processing)
- Identification of body fluids (saliva and semen)
- How to triage SAEs
 - Every case is different
 - Multiple suspect / consensual partner cases
 - Male victim cases
 - Female suspect cases
- What to take forward
 - STR testing
 - Y-STR testing
 - Both



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“Direct to DNA” Approach

- Initial processing includes SAE kit orifice swabs:
 - Vaginal and anal
 - Oral (depending on history)
 - Also most relevant swabs (i.e. thighs, neck, breasts, etc.)
- Differential extractions performed on vaginal/anal, thighs
- No initial semen screening performed on SAE kit items
- Subsequent re-processing of additional swabs and/or clothing may be needed
- Y-screening [Auto]/[Y]
 - <10 STR
 - 10-40 STR and/or Y-STR
 - >40 Y-STR
 - ≤0.001ng/μl total male DNA – stop at quant

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Considerations Over Time

- Analysts given more discretion over sample processing:
 - Oral swabs only on a case-by-case basis
 - Epithelial fractions not purified initially, only when needed
 - Only stain/search slides for samples that are amplified
 - Y-STR analysis may be more beneficial regardless of [Auto]/[Y] ratio (low quant value)
- Traditional semen screening on clothing & bedding
 - Touch on a case by case basis
- Amplification decision based on:
 - [Auto]/[Y] ratio ranges
 - Total human and male quant values

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Where do we go from here?

- Processing a lot of differential samples up front
 - No initial screening, so must treat anything with potential semen as a differential
 - Very time-consuming, waste of reagents/consumables
- Multiple rounds of processing
 - "Triage" samples based on medical report/case synopsis
 - Not everything from SAEK processed initially
- Many sperm slides very "dirty"
 - Hard to view sperm
 - Tried different slides and dyes: didn't really help



Introduction

- Promega Casework Direct Kit, Custom
 - Used for rapid processing of swabs from casework samples prior to quantification
 - Used as a male screening tool for sexual assault swabs/cuttings
 - No subsequent purification of the Prototype Casework Direct lysate is required prior to STR amplification
 - Kit contents:
 - Casework Direct Reagent
 - 1-Thioglycerol (as a reducing agent)
 - 5X AmpSolution™ Reagent- must be added to Plexor® HY quantifications
 - Casework (CW) tubes and spin baskets recommended for this procedure to eliminate the need for tube transfers*
 - * Issue with tubes not sealing very well without the spin basket; decided against using them for Casework Direct processing; currently use QIAGEN Lyse + Spin 2mL tubes



Introduction

- | | |
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| <ul style="list-style-type: none"> • Casework Direct <ul style="list-style-type: none"> - Simple <ul style="list-style-type: none"> • 2-step procedure - Fast <ul style="list-style-type: none"> • 30 min incubation • 5 minute centrifuge spin - No wash steps - Minimal potential for DNA loss - Use lysate for both quantification and amplification - ~\$6.40 per sample | <ul style="list-style-type: none"> • EZ1 <ul style="list-style-type: none"> - Preprocessing steps - 1-2 hour incubation times - Cartridge-based purification - Additional machine run time - Multiple wash steps - Multiple transfer steps - Potential for DNA loss <ul style="list-style-type: none"> • Binding efficiency of DNA to resin • Washes • Elution efficiency - ~\$9.90 per sample |
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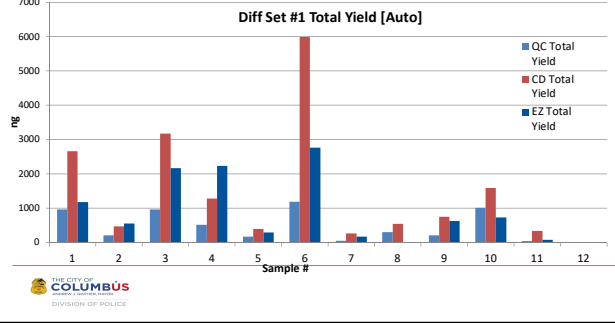


Methods – Differential & Blood

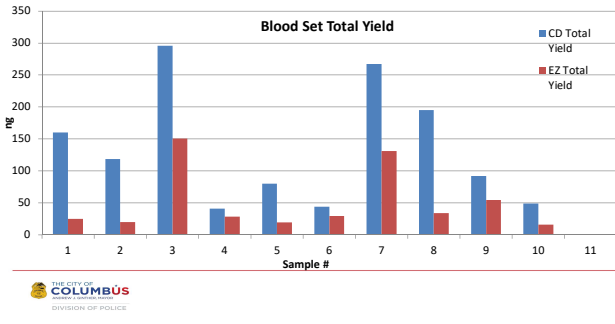
- Differential and blood samples were extracted using the QIAcube, Casework Direct, and current extraction (EZ1) methods
- Extractions were performed following the provided manufacturer protocols, with a few minor variations
 - Recommended 100-400µl – validated 300µl for Casework Direct*
 - *300µl determined to be best volume for amount of swab extracted
- One swab per sample was used for each differential sample and approximately 3mm² cuttings per blood sample were used
- All extracted samples were quantified with Plexor® HY
 - Recommended addition of 5X AmpSolution™ Reagent to Reaction Mix
- PowerPlex® Fusion 6C amplification
- 3500 capillary electrophoresis
- GeneMapper™ ID-X v1.4

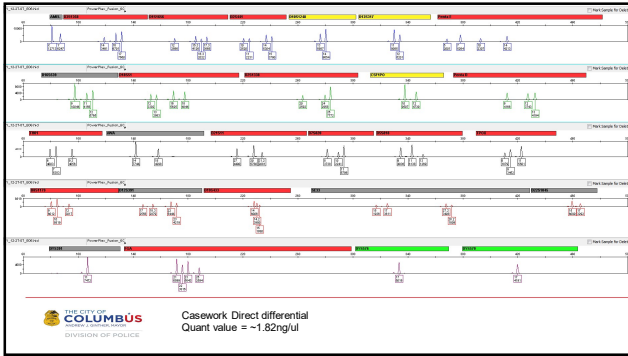


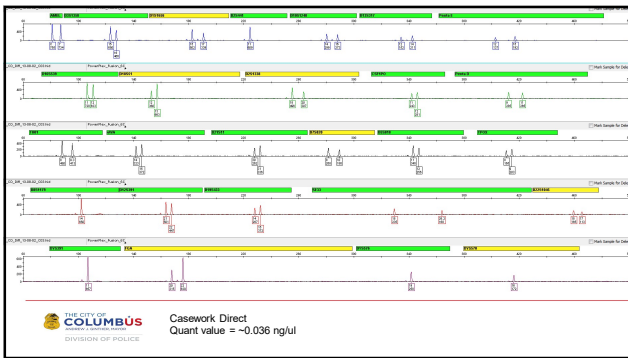
Results

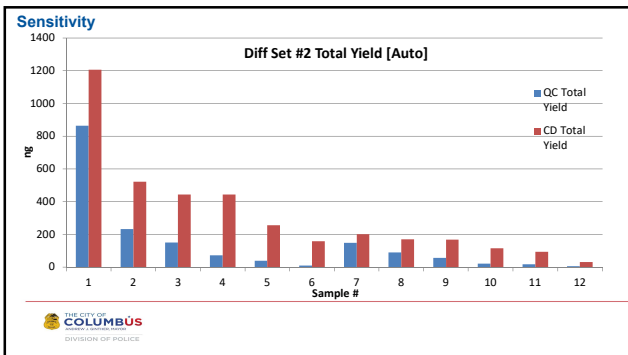


Results









Methods: Trace Samples

- 30 minute incubation vs. 60 minute incubation
- 70°C vs 80°C incubation temperature
- Addition of 250µl and 200µl vs 300µl of CWD reagent
- No significant difference in DNA recovery
 - CWD extract too dilute and microconning not recommended
 - EZ1 still the preferable extraction method for low-level samples at this time



Methods: Cigarette Butts

Samples	EZ1	CWD
	[Auto] ng/µL	[Auto] ng/µL
Cigbutt_1	0.2748	0.8771
Cigbutt_2	0.0211	0.0186
Cigbutt_3	0.4323	0.1948
Cigbutt_4	0.0002	N/A
Cigbutt_5	0.1366	0.0343

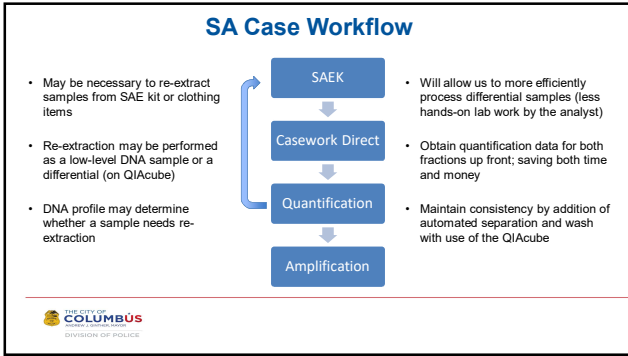
- No significant difference in quant values for cigarette butts extracted using CWD vs. EZ1
- More dropout observed in F6C CWD profiles when amplified versus EZ1
- Overall, CWD extracts too dilute and currently no method to concentrate samples



Considerations, Casework Direct

- High-level DNA samples will continue to extract over time, may need to re-quant if leaving stored in fridge for an extended period of time before amplification
- Consumption samples will be extracted normally, not with Casework Direct
- For lower level samples it may be necessary to perform an additional extraction to maximize the DNA yields, but the benefits of being able to quickly obtain Y-screening results from sexual assault samples will outweigh the disadvantage of not always being able to proceed directly to amplification with some samples
 - Based on more recent casework data, CWD typically yields higher quantification values
 - Usually not beneficial to re-extract on EZ1 unless as a differential





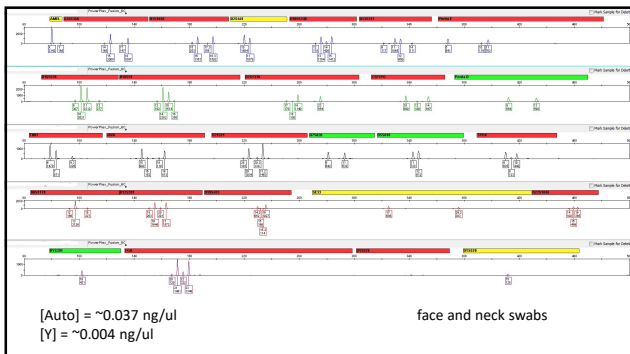
CWD Y-Screen Case Example

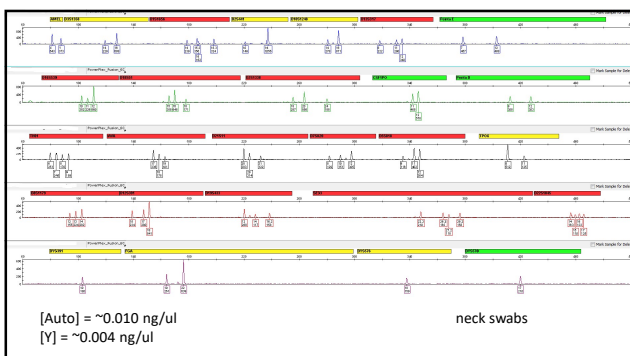
Sample	[Auto]	[Y]	[Auto]/[Y]
CWD vaginal	178.0050	0.0732	2433.1418
EF Diff vaginal	28.8832	0.0022	12877.5738
SF Diff vaginal	0.6540	0.1351	4.8423
CWD anal	0.0787	0.0111	7.1203
CWD oral	5.1713	N/A	N/A
CWD external genital	11.2217	0.0182	615.4170
EF Diff external genital	18.1827	0.0085	2126.6474
SF Diff external genital	0.1768	0.0660	2.6770
CWD inside of jeans	0.0087	0.0010	8.8902
CWD lips/face	0.0085	N/A	N/A
CWD neck	0.0485	0.0133	3.6539

Impacts on Casework

- No "limit" on sample number for initial Casework Direct extraction
 - Process samples faster and reduce TAT
- Very few samples require a differential extraction
 - Validation of STRMIX® has helped for interpretation and reporting of mixture samples
 - Use of differential extractions for samples with high [Auto]/[Y] ratio (~>40)
- No real improvement of "touch" SA samples upon re-extraction on the EZ1
- Better resolution of minor F6C alleles with higher [Auto]/[Y] ratios (range of 30-40)
- Lower [Y] yielding more interpretable STR profiles
 - F6C validation showed loss of minor male @ ~0.008 ng/ul [Y]
 - Able to obtain interpretable male profiles @ ~0.004 ng/ul [Y]

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Conclusions, Casework Direct

- The Casework Direct extraction method is a robust extraction method that can be used in place of our current differential extraction method as a Y-screening tool
- It enables us to obtain quantification data from both the epithelial and sperm cell fractions in one short extraction process
- Furthermore, it is often possible to proceed with amplification and analysis which provides additional time and cost savings
- The Casework Direct extraction method is also a worthwhile alternative to our current high-level DNA samples - the shortened extraction time enables us to more effectively analyze batches of these sample types
- Results from low-level DNA samples do not support use of Casework Direct as a final extraction method – looking at other alternative methods



Looking into the future...

- Alternative "touch" extraction method currently being evaluated
- Preliminary data with Promega's Maxwell® Extraction Robot shows better recovery of DNA versus current EZ1 method
- Can accommodate a larger number of samples per robot and requires less processing time



Looking into the future...

- Post-PCR clean up method is currently being validated using Qiagen's QIAquick® PCR purification kit
- Used to remove impurities introduced during amplification that can affect subsequent downstream applications
- Looking to also try this "clean-up" method with Casework Direct extracts both pre- and post-PCR



Thank You!

- Promega
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 - Emma Becker
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 - Mandy Fashano
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 - Hope Olson
 - Bob Parker



QUESTIONS?



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